Neurobiology of Lithium: An Update

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Lithium remains a first-line approach for the treatment of acute mania and the prophylactic management of manic-depressive illness, yet the underlying neurobiological mechanisms remain as yet undefined. In this paper we critically examine the accumulated preclinical and clinical evidence for the action of lithium in the brain and suggest areas that may be most productive for future investigation, i.e., membrane transport systems, neurotransmitter receptor regulation, second messenger generating systems, protein kinase C (PKC) regulation, and gene expression. In their experimental design, preclinical investigations have often jeopardized the physiologic relevance of their studies by a relative lack of attention to issues such as therapeutic concentrations, acute versus chronic exposure, and a lack of adequate cation and/or psychotropic controls. Future studies should account for the established prophylactic efficacy of lithium, the higher risk for relapse into mania after abrupt discontinuation, the ability of lithium to stabilize recurrent depression associated with unipolar disorder, and the efficacy of lithium in the treatment of refractory major depressive disorder in the presence of an antidepressant. Studies of the action of lithium in receptor mediated phosphoinositide signaling in the brain over the past several years have opened up heuristic lines of investigation that stem from lithium's uncompetitive inhibition of the enzyme inositol monophosphatase. Subsequent studies involving regulation of inositol transport, PKC isozymes and activity, and the expression of the major PKC substrate MARCKS (myristoylated alanine-rich C-kinase substrate) have offered potential avenues for understanding the complexity of the action of long-term lithium in the brain. These studies will offer us a better understanding of the neuroanatomical sites of action of lithium and together with ongoing clinical investigations using brain imaging in patients with manic-depressive illness a more complete understanding of the pathophysiology of this disease.

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Within decades of its discovery, lithium was found to be the most soluble salt of uric acid and early observations of Alexander Ure in the 1840s addressed the potential medicinal properties of lithium in gout. Sir Alexander Garrod was the first to introduce the oral use of lithia salts as a treatment for gout or "uric acid diathesis," which was believed to encompass affective symptoms of both mania and depression.^{1–3} However, it wasn't until the observations of the American physician John Aulde and the

Danish internist Carl Lange in the 1880s that lithium was considered to be a treatment of recurrent symptoms associated with depression independent of gout.^{4,5} After falling into disrepute as a medication because of serious toxicity associated with its widespread use in elixirs and tonics as well as a salt substitute, it was the rediscovery by Cade 48 years ago and seminal clinical studies by Schou in the 1950s that positioned lithium as an effective antimanic treatment and prophylactic therapy for manic-depressive illness.^{6,7} Since Cade's observations in 1949, numerous studies have explored potential mechanisms for the therapeutic action of lithium in the treatment of manic-depressive illness. Early studies focused on lithium's property of being a monovalent cation, its role in sodium transport, and its effect on electrophysiologic properties of cells. As neurotransmitter systems in brain were discovered over the past 45 years, the effects of lithium on these systems and their receptors have been examined in both animal and clinical investigations. Moreover, it has become increasingly apparent that the underlying pathophysiology of manic-depressive illness derives from a dysregulation of multiple signaling pathways in limbic and limbic-related regions of the brain, which results in recurrent clinical affective symptomatology.⁸⁻¹¹ Unfortunately, the majority of these studies have suffered from a number of limitations, and interpretation has been confounded by several issues including a lack of attention in

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experimental design to acute versus chronic effects as well as to clinically relevant lithium concentrations; a lack of specificity of pharmacologic agonists and antagonists; examination of one neurotransmitter system to the exclusion of others; overinterpretation in clinical studies of the relevance of peripheral findings to the CNS; and often small effects that have been difficult to replicate. In this review, we will focus upon several areas of research that have provided a strategically important direction or hold the promise of a new perspective for understanding the therapeutic action of lithium in the brain.

Several points regarding the clinical action of lithium in the treatment of manic-depressive illness are worthy of note as related to its mechanism of action in the brain. It is generally agreed that the action of lithium that is most efficacious in the treatment of manic-depressive illness is its prophylactic efficacy in preventing the recurrence of affective episodes and stabilizing the course of the illness during long-term treatment.¹² Upon even rapid discontinuation of lithium in patients who have undergone chronic treatment for many years, symptomatology does not immediately return, but there appears to be a significantly increased risk for relapse over the ensuing months, highly suggestive of a withdrawal-like physiologic response.13-15 The observation that there is a predisposition during the withdrawal period to relapse into a manic episode should be of considerable interest to investigators. Lithium is also effective in the treatment of acute mania, but its action occurs over the course of 1 week to 10 days. In the treatment of unipolar illness, lithium has been shown to possess therapeutic benefit in the prevention of recurrent episodes of major depressive disorder and in the treatment of refractory episodes of major depressive disorder. In the latter case, the effect of lithium appears to occur in the presence of an already existing antidepressant and is apparent over a period of days to weeks, often at plasma concentrations that are significantly less than those used in the treatment of manic-depressive illness.9,12 As we examine the mechanism of action of lithium in the brain, it will be important to note that while it is highly likely that more than one molecular target in the brain accounts for these clinical properties, common sites of action may trigger events that result in both enhancement of antidepressant activity as well as stabilization of dysregulation in multiple signaling pathways in the brain. Moreover, such sites of action for lithium should account in some way for its specificity of action at therapeutic concentrations in critical regions of the brain.

LITHIUM AND MEMBRANE TRANSPORT

Both membrane transport systems and ion channels play roles in the regulation of intracellular lithium. Transport systems may be driven by either ATP (adenosine triphosphate), e.g., the Na,K-ATPase, or by the net free energy of transmembrane concentration gradients, e.g., the sodiumcalcium exchanger. These transport systems are likely to be crucial for the regulation of resting lithium in the bulk cytoplasm, as they essentially regulate all steady-state intracellular ion concentration. While membrane transport systems may exist that specifically recognize lithium and regulate its transmembrane concentration, e.g., a gradientdependent sodium-lithium exchange process,^{16,17} it is arguably more likely that the primary regulation of lithium is affected by transport systems that accept the lithium ion as a substitute for their normal ionic substrates. While clinical studies over the years have been constrained by relatively small and often variable findings, there is evidence that Na,K-ATPase activity may be reduced especially in the depressed phase of both unipolar depression and manic-depressive illness, which is associated with an increase in sodium retention (see reviews in references 9 and 18). Furthermore, chronic lithium treatment has been observed to result in an increased accumulation of lithium and activity of Na,K-ATPase in erythrocyte membranes and concomitant reduction of intraerythrocyte sodium and calcium in patients with manic-depressive illness.9 Since free calcium ion concentration tends to parallel free sodium concentration, this may account for observations that intracellular calcium is increased in patients with manicdepressive illness.¹⁹ These data related to Na,K-ATPase should be viewed with caution, however, since recent data also support the evolution of specific gene products expressed and posttranslationally regulated and unique not only to neurons but among brain regions.²⁰⁻²⁵ Thus, while the erythrocyte may be used as a peripheral model for lithium transport, extrapolations to lithium homeostasis in the brain or as a potential genetic model for variations in ionic homeostatic processes in the brain underlying the pathophysiology of a disease such as manic-depressive illness remain highly speculative. Since most data seem to indicate that lithium achieves

an approximately equal distribution across the plasma membrane barrier while leading to only small depolarizations, it should be appreciated that the predominant form of lithium transport must be efflux to oppose any resting conductances that would tend to concentrate positively charged ions within the negatively charged intracellular milieu, which is reflected in reported brain; serum ratios in the range of 0.76.²⁶ While it is the balance of resting lithium conductance and net transport-efflux mechanisms that regulate steady-state lithium homeostasis, the gating of ion channels on the time scale of the channel activity will alter this homeostasis to varying degrees. Thus, activation of voltage-dependent sodium channels may play a significant role in the regulation of intracellular lithium concentration in neurons, such that lithium concentration might be elevated in active neurons, on the basis of the influx of lithium through sodium channels. However, simple calculation reveals that the flux of lithium through voltage-dependent sodium channels is unlikely to be important in the regulation of lithium homeostasis, even on a short-term basis or as a consequence of high levels of neuronal activity. Moreover, in the local environment of a dendritic spine, the surface area to volume ratio becomes relatively large, such that the lithium component of a synaptic current can cause very significant increases in the local lithium concentration—as much as a fivefold to tenfold increase in intracellular lithium following a train of synaptic stimuli.²⁷ Such an activity-dependent mechanism for creating focal increases of intracellular lithium at sites of high synaptic activity may be crucial for lithium's therapeutic specificity and ability to regulate synaptic function in the brain.

LITHIUM AND NEUROTRANSMITTER AND NEUROPEPTIDE SYSTEMS

Over the past 30 years, clinical and preclinical studies have focused upon the effects of both acute and chronic lithium on the regulation of monoamine neurotransmitter systems in brain; modest attention has been given to amino acid and neuropeptide neurotransmitters (reviewed in references 9, 12, 28, and 29). More recently attention has turned to lithium-induced changes in second messenger systems and gene expression, which have served to form heuristic lines of investigation actively being pursued.

Norepinephrine

Findings in clinical studies of lithium treatment have generally been confounded by changes in affective state and associated changes in activity level, arousal, and sympathetic outflow and have provided conflicting evidence for an effect of lithium on norepinephrine levels and turnover in brain. On the other hand, preclinical studies appear to support an action of acute lithium in reducing the β -adrenergic stimulated AC (adenylyl cyclase) response, and chronic lithium in facilitating the release of norepinephrine, possibly via effects on the presynaptic α_2 autoreceptor, as well as blockade of the β -adrenergic receptor supersensitivity after presynaptic depletion of norepinephrine (see review in reference 9).

Dopamine

Clinical studies of dopamine metabolites in the periphery of bipolar patients have similarly been confounded by mood and activity state. Chronic lithium has been reported to prevent haloperidol-induced dopamine receptor up-regulation and induce supersensitivity to iontophoretically applied dopamine or intravenous apomorphine.^{30–33} Interestingly, a number of studies have reported a lack of effect if lithium is administered after the induction of dopamine supersensitivity suggesting that in this model, lithium exerts its greatest effects prophylactically.^{34–37} Lithium also appears to block amphetamine-induced behavioral changes in both animals and humans.^{28,29,35,36,38–41}

Serotonin

Preclinical studies indicate that the effects of lithium on serotonin (5-HT) function may occur at multiple levels and result in an enhancement of serotonergic neurotransmission, although its effects on 5-HT appear to vary depending on brain region, length of treatment, and 5-HT receptor subtype (see review in reference 9). Interpretation of studies attempting to clarify the roles of presynaptic versus postsynaptic receptors in mediating the effects of lithium on 5-HT function have been confounded by the relative lack of understanding of the numerous receptor subtypes and their distribution, as well as the existence of subtype-specific agonists and antagonists. However, there is accumulating evidence that lithium produces a subsensitivity of presynaptic inhibitory 5-HT_{1A} receptors, which can result in a net increase of the amount of 5-HT released per impulse.⁴²⁻⁴⁷ These findings are consistent with observations that short-term lithium enhances the efficacy of the ascending (presynaptic) 5-HT system^{48,49} and have formed the basis for a series of clinical investigations demonstrating the efficacy of lithium as an adjunct to antidepressants in the treatment of refractory depression.

Acetylcholine

Neurochemical, behavioral, and physiologic studies have all suggested that the cholinergic system is involved in affective illness⁵⁰ and that lithium enhances the synaptic processing of acetylcholine (ACh) in rat brain. Notably, lithium effectively potentiates seizures induced by a muscarinic agonist which are markedly attenuated by central myo-inositol administration,^{51,52} consistent with effects of lithium on receptor-mediated PI (phosphoinositide) signaling discussed below. In addition, studies by Evans and colleagues⁵³ have suggested that the role of lithium in the lithium-pilocarpine seizure model appears to occur through a presynaptic facilitation of excitatory neurotransmission mediated by protein kinase C (PKC). Thus, lithium may target cholinergic neurotransmission and chronically may play a role in preventing muscarinic receptor supersensitivity through interaction within PI signaling systems. These findings are consistent with an effect of chronic lithium in preventing receptor-mediated supersensitivity as observed in both the dopaminergic and noradrenergic systems and suggest a putative site of action for lithium beyond the receptor at receptor-effector coupling and/or intracellular second messenger systems.

Amino Acids

While relatively less attention has been paid to effects of lithium on amino acid and neuropeptide regulation in brain, dysregulation of GABAergic neurotransmission (gamma-aminobutyric acid [GABA] is the major inhibitory neurotransmitter in brain) has been postulated to play a role in the etiology of affective disorders.^{54–56} In an albeit limited series of patients, GABA has been observed to be re-

duced in CSF of depressed patients⁵⁷⁻⁵⁹ and in the plasma of bipolar patients.⁶⁰ Previously low levels of plasma and CSF GABA appear to normalize in bipolar patients being treated with lithium,^{61,62} though baseline GABA levels appear unrelated to clinical responsiveness to lithium.⁶³ While preclinical studies have been constrained by the limitations in design and methodology noted earlier, investigators have reported that lithium produced elevations in GABA in the striatum and midbrain,64-66 GABA turnover in the hippocampus and striatum,67 and a potentiation of kainic acid-evoked [³H]GABA release in striatal neurons.⁶⁸ The effect of lithium on glutamate, a major excitatory neurotransmitter in brain, has been studied more recently by using monkey cerebral cortical slices-lithium was found to stimulate glutamate release at doses ranging from 1.5 to 25 mM.⁶⁹ It is of interest that the proconvulsant action of lithium in pilocarpine-treated rats described earlier is blocked by the noncompetitive NMDA (N-methol-p-aspartate) receptor antagonist, MK-801.70 However, chronic in vivo lithium treatment results in a transient impairment of ACh-induced potentiation of current responses elicited by NMDA in hippocampal CA1 neurons.71

Neuropeptides

Preclinical studies of neuropeptides have revealed diverse effects of lithium on multiple systems including the opioid peptides, substance P, tachykinin, neuropeptide Y, neurokinin A, and calcitonin gene-related peptide. Comparisons between studies are difficult due to differences in time and dose of lithium administration; however, for the most part, an increase in neuropeptide levels in brain has been observed and in the case of both dynorphin and tachykinin, the increase is associated with an increase in mRNA.^{72,73} It has also been reported that chronic lithium abolishes both the secondary reinforcing effects of morphine and the aversive effects of the opioid antagonist, naloxone.^{74–78} However, in one of the few applicable clinical studies, CSF levels of various pro-opiomelanocortin peptides were examined in euthymic bipolar patients before and during lithium treatment; no significant effects of lithium on the CSF levels of any of the peptides were observed.^{79,80} Alterations in the regulation of these neuropeptides, particularly in basal ganglia, may be of interest in regard to the commonly observed lithium-induced sideeffect of tremor.

LITHIUM AND SIGNAL TRANSDUCTION

Phosphoinositide Cycle

Lithium, at the rapeutically relevant concentrations in brain, is a potent inhibitor of the intracellular enzyme, inositol monophosphatase (K_i = 0.8 mM), which plays a major role in the recycling of inositol phosphates.^{81,82} Since the brain has limited access to inositol other than that derived from recycling of inositol phosphates, the ability of a cell to maintain sufficient supplies of myo-inositol can be crucial to the resynthesis of the PIs and the maintenance and efficiency of signaling.⁸³⁻⁸⁵ Furthermore, since the mode of enzyme inhibition is uncompetitive,^{86,87} the effects of lithium have been postulated to be most pronounced in systems undergoing the highest rate of PIP₂ (phosphatidylinositol 4,5-bisphosphate) hydrolysis. Thus, Berridge and associates^{88,89} first proposed that the physiologic consequence of lithium's action is derived through a depletion of free inositol, and that the selectivity of lithium could be attributed its preferential action on the most overactive receptor-mediated neuronal pathways. Since several subtypes of adrenergic, cholinergic, and serotonergic receptors are coupled to PIP₂ turnover in the CNS, such an hypothesis offers a plausible explanation for the therapeutic efficacy of lithium in treating the dysregulation of neurotransmitter signaling as underlying the pathophysiology of bipolar illness.^{8,10} The preponderance of data indicates that the effects of lithium on PI signaling can be prevented and reversed in the presence of high concentrations of exogenous myo-inositol. 52,84,90-92 In a recent preliminary clinical investigation that used proton magnetic resonance spectroscopy (1H-MRS), lithium administration to bipolar depressed and manic patients produced a significant reduction in the levels of myo-inositol in frontal (but not occipital) cortex; these changes are observed after 5 days of treatment and persist for at least 3 to 4 weeks.⁹³ The action of lithium in inhibiting the recycling of inositol through the receptor-mediated hydrolysis of PIP₂ is thought to cause an accumulation of critical pools of diacylglycerol resulting in activation of PKC isozymes responsible for mediating long-term alterations in cell function.^{8,10,94} In fact, downstream effects of chronic lithium on embryonic development and regulation of PKC and MARCKS protein triggered by changes in PI signaling have also been shown to be dependent upon myo-inositol availability.95-97 Studies wherein the effects of myoinositol have been difficult to demonstrate may be attributable, at least in part, to significant differences in inositol transport and accumulation among cell types.⁸⁵ Thus, overall, both the preclinical and clinical data suggest that although lithium does bring about a relative depletion of *myo*-inositol in the brain, its therapeutic effects are likely mediated by a secondary cascade of signaling changes, rather than reductions in *myo*-inositol per se.^{9,10,98}

Adenylyl Cyclase

The other major receptor-coupled second messenger system in which lithium has been shown to have significant effects is adenylyl cyclase (AC), which generates cyclic adenosine monophosphate (cAMP). cAMP accumulation by various neurotransmitters and hormones is reported to be inhibited by lithium at high therapeutic concentrations and above both in vivo and in vitro, but the sensitivity, especially in brain, appears to be less than that observed in the PI system.^{45,99–107} In fact, lithium inhibition of vasopressin-sensitive or thyroid-stimulating-hormonesensitive AC is generally believed to underlie two of lithium's more common side effects, namely nephrogenic diabetes insipidus and hypothyroidism.¹⁰⁸⁻¹¹¹ Norepinephrine- and adenosine-stimulated cAMP accumulation in rat cortical slices are inhibited significantly by 1 to 2 mM of lithium; in human brain tissue, the IC_{50} for lithium inhibition of norepinephrine-stimulated cAMP accumulation is approximately 5 mM.¹¹² Prolonged lithium exposure produces little effect on β-adrenergic-stimulated AC but an increase in basal cAMP has been observed.^{100,107,113} Data suggest that lithium's inhibition of AC in vitro may be due to competition with Mg⁺⁺ for a binding site on the catalytic unit of AC.99,107 However, the inhibitory effects of chronic lithium treatment on rat brain AC are not reversed by Mg⁺⁺, and these effects still persist after washing of the membranes but are reversed by increasing concentrations of GTP (guanosine triphosphate).¹⁰⁴ These results suggest that the physiologically relevant effects of chronic lithium may be exerted at the level of signal-transducing G proteins at a GTP-responsive step.

G Proteins

Since lithium has been shown to affect both PI turnover. and AC activity, a series of studies have focused on mechanisms shared by these two major second messenger generating systems, namely the signal-transducing G proteins. As noted above, experimental evidence has shown that lithium may alter receptor coupling to PI turnover in the absence of consistent changes in the density of the receptor sites themselves. When fluoride ion or GTP analogs that directly activate G protein-coupled second messenger responses were used, studies in rat brain after chronic lithium exposure have revealed either no change¹¹⁴ or an attenuation¹¹⁵ of PI turnover. While there have been reports of a direct effect of lithium on receptor-mediated G protein binding of guanine nucleotides in brain membranes, such data have been difficult to replicate and are physiologically inconsistent.116-118

Investigations have also addressed the role of G proteins in the action of lithium-induced attenuation of receptor-mediated AC activity in both rodents and humans. These studies have revealed evidence for an increase by chronic lithium of pertussis toxin-catalyzed [³²P]ADPribosylation of inhibitory G proteins in both rat brain as well as platelets from volunteer subjects treated for 2 weeks.^{119,120} These data provide no evidence for a change in the amount of the G proteins and suggest a stabilization of the inactive undissociated $\alpha\beta\gamma$ heterotrimeric form of the inhibitory G protein. This was accompanied by an enhancement of both basal and postreceptor-stimulated AC in both the rat brain and platelet preparations, which is consistent with an uncoupling of the tonic inhibitory influence of the G protein.¹¹ These data are also consistent with

the studies demonstrating that effects of chronic lithium exposure may be exerted at a GTP-responsive step and may result in an alteration in the conformational state (active/inactive) of the G protein. Such a contention is supported by the recent observation that chronic in vivo lithium administration reduces the subsequent in vitro sensitivity of rat cortical membranes to guanine nucleotideinduced reductions in pertussis toxin catalyzed [32P]ADPribosylation.¹²¹ This is also consistent with the results of Mork and Geisler,¹⁰⁵ who demonstrated that the addition of exogenous guanine nucleotides (but not Mg⁺⁺) was able to overcome the effects of lithium on rat brain AC activity. Most recently, it has been shown that lithium promotes calpain-mediated proteolytic cleavage of G_o, effects which have been postulated to occur via a stabilization of the G protein in its $\alpha\beta\gamma$ heterotrimeric conformation¹²²; moreover, these investigators also found that the addition of GTP γ S was able to overcome the effects of lithium. Thus, these findings suggest that chronic lithium may lead to the GTP-dependent stabilization of G_i (and G_o) in the undissociated $\alpha\beta\gamma$ conformation.

At present, the possible effects of chronic lithium on the absolute levels of Gq and Gq remain unclear; two independent laboratories have not observed any alterations,^{120,123,124} whereas another laboratory has reported small but significant decreases in the levels of the Gq, $G\alpha_{11}$, and $G\alpha_{12}$ in rat frontal cortex.¹²⁵ However, chronic lithium administration appears to reduce the mRNA levels of a number of G proteins in rat brain, including α_i , α_i , and α_2 .^{123,125} This is of interest in light of studies implicating PKC in the down-regulation of G protein mRNAs,¹²⁶ and its posited role in the action of chronic lithium (see below). In summary, although there is evidence that competition with magnesium accounts for some of the in vitro effects of lithium on G proteins and speculation that an interaction with GTP binding might be relevant to the chronic effects of lithium, a direct effect of lithium on guanine nucleotide activation of G protein remains unsubstantiated. Most recently, investigators have demonstrated that lithium alters the levels of endogenous ADP-ribosylation in C6 glioma cells¹²⁷ and in rat brain,¹²⁸ suggesting another mechanism through which chronic lithium may indirectly regulate the activity of these critical signaling proteins. Thus, overall the data suggest that the long-term effects of chronic lithium on G protein may more likely be attributable to an indirect posttranslational modification of the G protein(s) and a relative change in the dynamic equilibrium of the active/inactive states of protein conformation, potentially resulting in modulation of receptor-mediated signaling in critical regions of the brain.

Protein Kinase C and Phosphoprotein Substrates

As noted above, recent evidence accumulating from various laboratories points to a role for PKC in mediating the action of lithium in a number of cell systems and the

brain (see reviews in references 10 and 93). PKC is a family of at least 12 phosphorylating isozymes with differing tissue, intracellular and regional distribution within the brain, second messenger activators, and substrate affinities, indicating distinct cellular functions.¹²⁹⁻¹³² Posttranslational phosphorylation of selective PKC protein substrates within the cell are responsible for regulation of processes critical to cell secretion, membrane trafficking, transcription, ion transport, receptor signaling, and transformation. Recent studies have demonstrated that chronic lithium administration in rats results in a significant decrease in the membrane-associated PKC α and ϵ isoforms in the hippocampus.^{133,134} It is noteworthy that exposure of neuroblastoma cells¹³⁵ or PC12 cells¹³⁶ to 1 mM of lithium in vitro produces isozyme-selective decreases in PKC α and (in the case of PC12 cells) PKCE. Recent studies have also investigated the effects of valproate (an anticonvulsant with demonstrated antimanic properties) on PKC isozymes and substrates. Chronic valproate has been found to produce strikingly similar reductions in the levels of PKC α and ε .¹³⁷ These findings are of particular interest and have led to a pilot clinical investigation showing efficacy of the use of tamoxifen, an effective PKC inhibitor, in the treatment of acute mania.¹²¹ The precise mechanism(s) by which chronic lithium treatment produces these isozyme-specific effects is presently unclear; however, PKC subspecies are known to exhibit subtle differences in their biochemical characteristics and cellular localization and also differ in their susceptibility to degradation after activation.¹³⁸⁻¹⁴⁰ In addition, the down-regulation of the ε PKC isoform after chronic lithium exposure can be reversed by the coadministration of myo-inositol in vivo,¹³⁴ a finding that is consistent with the action of chronic lithium resulting in an inositol depletion, accumulation of DAG, (diacylglycerol) and subsequent activation of PKC isozymes as described above. While we as yet do not fully understand the precise profile of PKC isozymes activated by lithium, we have begun to investigate critical protein substrates for PKC that may provide further insight into the mechanism of longterm action of lithium in the brain.

The activation of PKC results in the phosphorylation of a number of membrane-associated phosphoprotein substrates, the most prominent of which in brain is the myristoylated alanine-rich C-kinase substrate (MARCKS). Direct activation of PKC by phorbol esters in immortalized hippocampal cells will effectively down-regulate the MARCKS protein.¹⁴¹ Chronic lithium administered at therapeutically relevant concentration (1 mEq/kg in brain) to rats over a 4-week period has been shown to result in a marked reduction in MARCKS in the hippocampus, which is not observed after acute treatment and persists beyond treatment discontinuation.¹⁴² The down-regulation of MARCKS expression in brain has also been observed at concentrations as low as 0.7 mM.¹³⁴ The lithium-induced reduction in MARCKS has recently been replicated in im-

mortalized hippocampal cells and shown to be prevented and reversible in the presence of elevated inositol concentrations.95 Furthermore, activation of muscarinic receptorcoupled PI signaling significantly potentiates the downregulation of MARCKS protein induced in the presence of 1 mM of lithium, which supports the role of the PI signaling pathway and PKC isozymes in this long-term action of lithium.95 MARCKS binds calmodulin in a calciumdependent fashion and crosslinks actin at the plasma membrane; both these events are inhibited by PKC-mediated phosphorylation that serves to translocate MARCKS from the membrane to the cytosol.^{143,144} MARCKS has been implicated in cellular processes associated with cytoskeletal restructuring and signaling that may be related to longterm neuroplastic changes in processes associated with receptor signal transduction and neurotransmitter release. Recent studies have indicated that this action of chronic lithium on MARCKS protein expression is not shared by psychotropic drugs in general, but is a property of valproate at therapeutic concentrations relevant to the treatment of acute mania.145,146 Thus MARCKS may represent a clinically relevant target for the mood-stabilizing action of chronic lithium, which serves to regulate aberrant signaling in the brain of patients suffering from manicdepressive illness.

LITHIUM AND GENE EXPRESSION

The prophylactic efficacy of lithium generally requires a number of weeks to develop,^{12,94,98} suggesting long-term neuroplastic alterations potentially mediated at the genomic level. Indeed, increasing evidence suggests that lithium affects gene expression, possibly via PKC-induced alterations in nuclear transcription regulatory factors responsible for modulating the expression of specific genes.^{147,148} Several recent studies have demonstrated that lithium alters the expression of the immediate early gene c-fos in different cell systems including the brain.94,149-152 These lithium-induced effects on the expression of c-fos mRNA, generally thought to represent a "master switch" to turn on a "second wave" of specific neuronal genes of functional importance, offer a mechanism for affecting long-term events in the brain. Chronic in vivo administration of lithium resulting in clinically relevant levels alters the expression of a number of genes in rat brain, several of which are known neuromodulatory peptide hormones (prodynorphin, preprotachykinin) and their receptors (glucocorticoid type II), and are known to contain PKC-responsive elements. 72,73,153-156 Recent studies have also demonstrated that the effects of both lithium and valproate on the regulation of AP-1 DNA binding activity in cultured cells^{157,158} may be mediated at least in part by PKC. The demonstration of the long-term modulation of the genetic expression of critical proteins involved offers new strategies for unraveling the complex physiologic effects of chronic lithium in the prophylaxis of recurrent episodes of affective illness in patients with manic-depressive illness.

NEUROANATOMICAL SITE OF ACTION

While data related to neuroanatomical localization of lesions in brains of patients who have manic-depressive illness are only now being accumulated by using structural and functional neuroimaging strategies, alterations in right hemisphere regions related to limbic and frontal association areas have been of particular interest (see review in reference 11). The premise that lithium exerts its therapeutic actions by acting at such specific neuroanatomical sites and/or their cells of projection is supported by several lines of evidence. First, atomic absorption spectrophotometric, radiographic dielectric track registration, and nuclear magnetic resonance studies indicate that lithium does not distribute evenly throughout the brain after either acute or chronic administration, but preferentially accumulates in forebrain diencephalon, e.g., hypothalamus, and telencephalon structures, e.g., caudate and hippocampus.¹⁵⁹⁻¹⁷⁰ Second, as noted previously, preclinical studies of the effects of lithium on neurotransmitter systems have revealed changes, particularly in the 5-HT system, that are specific to certain brain regions. Third, the relative regional and cellular distribution of overactive ligand-gated ion channels in the brains of patients who have manic-depressive illness may be important in dictating relative rates of lithium transport.²⁷ Fourth, studies assessing PI turnover after lithium administration indicate regional differences in inositol depletion and agonist-stimulated [3H]IP accumulation, primarily between forebrain structures and hindbrain structures.^{115,169,171-175} Moreover, the effects of lithium will be most apparent in cells not only where inositol is limiting, but also those undergoing the greatest activation of receptor-mediated PI hydrolysis.¹⁷⁶⁻¹⁷⁸ Finally, regional brain distribution of PKC isozymes and alterations in MARCKS expression after chronic lithium, e.g., hippocampus, may confer even further specificity of action.^{133,142} Collectively, these studies indicate that the longterm therapeutic action of lithium may indeed possess cell and regional brain specificity that underlies its prophylactic efficacy in the treatment of manic-depressive illness.

CONCLUSIONS

Lithium remains our most effective treatment for reducing the frequency and severity of recurrent affective episodes in classic manic-depressive illness, yet despite extensive research, the underlying biological basis for the therapeutic efficacy of this drug remains unknown. Lithium is a monovalent cation with complex physiologic and pharmacologic effects within the brain. By virtue of the ionic properties it shares with other important monovalent and divalent cations such as sodium, magnesium, and calcium, its transport into cells provides ready access to a host of intracellular enzymatic events affecting short- and long-term cell processes. Thus, it is apparent that both clinical and preclinical investigations of the effects of lithium would result in the "dirty" characteristics of its multiple sites of pharmacologic interaction. However, recent investigations may implicate a role for ligand-gated ion channels in determining relative specificity in cell/ regional transport in brain that may reflect aberrant signaling underlying the pathophysiology of manic-depressive illness. Moreover, such dysregulation of signaling may also serve to dictate cell/regional extent of inositol depletion in the presence of chronic lithium that triggers PKCmediated downstream changes in expression of proteins such as MARCKS and neuroplasticity that restabilize a dysregulated pattern of signaling in critical regions of the brain. To what extent chronic lithium alters the patterns of gene and protein expression through PKC dependent versus independent mechanisms is yet to be discovered. However, future studies should begin addressing the relevance of such changes to the therapeutic efficacy of this psychotropic agent. Experimental designs should incorporate the critical clinical variables that are known regarding the use of lithium in the treatment of manic-depressive illness. As we gain understanding of the susceptibility genes underlying manic-depressive illness and the development of appropriate mutant mouse models, such clinically relevant studies of the mechanism of action of lithium will be facilitated in the future.

Drug names: amphetamine (Benzadrine), haloperidol (Haldol and others), naloxone (Narcan), tamoxifen (Nolvadex).

REFERENCES

- Garrod AB. The Nature and Treatment of Gout and Rheumatic Gout. London, England: Walton and Maberly; 1859
- Trousseau A. Clinique Medicale de l'Hotel-Dieu de Paris. 3rd ed. Paris, France: J-B Balliere et Fils; 1868
- 3. Ure A. Researches on gout. Medical Times 1844/1845;11:145
- Aulde J. The use of lithium bromide in combination with solution of potassium citrate. Medical Bulletin (Philadelphia) 1887;9:35–39, 69–72, 228–233
- Lange C. Om Periodiske Depressionstilstande og deres Patogenese. Copenhagen, Denmark: Jacob Lunds Forlag; 1886
- Cade JFJ. Lithium salts in the treatment of psychotic excitement. Med J Aust 1949;36:349–352
- Schou M. Lithium research at the Psychopharmacology Research Unit, Risskov, Denmark: a historical account. In: Schou M, Stromgren E, eds. Origin, Prevention and Treatment of Affective Disorders. London, England: Academic Press; 1979:1–8
- Lenox RH. Role of receptor coupling to phosphoinositide metabolism in the therapeutic action of lithium. In: Erlich YH, ed. Molecular Mechanisms of Neuronal Responsiveness. Adv Exp Biol Med, vol 221. New York, NY: Plenum Press 1987;515–530
- Lenox RH, Manji HK. Lithium. In: Nemeroff C, Schatzberg, eds. American Psychiatric Association Textbook of Psychopharmacology. Washington, DC: American Psychiatric Press; 1995;303–349
- Manji HK, Potter WZ, Lenox RH. Signal transduction pathways: molecular targets for lithium's actions. Arch Gen Psychiatry 1995;52:531–543
- 11. Manji HK, McNamara RK, Lenox RH. Mechanisms of action of lithium in bipolar illness. In: Montgomery S, Halbreich U, eds. Pharmacology of

Mood and Cognition. Washington, DC: American Psychiatric Press. In press

- Goodwin FK, Jamison KR. Manic-Depressive Illness. New York, NY: Oxford University Press; 1990
- Baldessarini RJ, Tondo L, Faedda GL, et al. Effects of the rate of discontinuing lithium maintenance treatment in bipolar disorders. J Clin Psychiatry 1996;57:441–448
- Faedda GL, Tondo L, Baldessarini RJ, et al. Outcome after rapid vs gradual discontinuation of lithium treatment in bipolar mood disorders. Arch Gen Psychiatry 1993;50:448–455
- Suppes T, Baldessarini RJ, Faedda GL, et al. Risk of recurrence following discontinuation of lithium treatment in bipolar disorder. Arch Gen Psychiatry 1991;48:1082–1088
- Hitzemann R, Mark C, Hirschowitz J, et al. RBC lithium transport in the psychoses. Biol Psychiatry 1989;25:296–304
- Sarkadi B, Alifimoff JK, Gunn RB, et al. Kinetics and stoichiometry of Nadependent Li transport in human red blood cells. J Gen Physiol 1978;72: 249–265
- El-Mallakh RS. Lithium Actions and Mechanisms. Washington, DC: American Psychiatric Press; 1996
- Dubovsky SL. Calcium channel antagonists as novel agents for manic-depressive disorder. In: Nemeroff C, Schatzberg A, eds. American Psychiatric Association Textbook of Psychopharmacology. Washington, DC: American Psychiatric Press; 1995;377–390
- Arystarkhoua E, Sweadner KJ. Isoform-specific monoclonal antibodies to Na,K-ATPase α subunits: evidence for a tissue-specific post-translational modification of the α subunit. J Biol Chem 1996;271:23407–23417
- 21. Grillo C, Piroli G, Gonzalez SL, et al. Glucocorticoid regulation of mRNA encoding (Na+K) ATPase α 3 and β 1 subunits in rat brain measured by in situ hybridization. Brain Res 1994;657:83–91
- 22. Lecuona E, Luquin S, Avila J, et al. Expression of the β 1 and β 2 (AMOG) subunits of the Na,K-ATPase in neural tissues: cellular and developmental distribution patterns. Brain Res Bull 1996;40:167–174
- Malik N, Canfield VA, Beckers MC, et al. Identification of the mammalian Na,K-ATPase subunit. J Biol Chem 1996;271:22754–22758
- Munzer JS, Daly SE, Jewell-Motz EA, et al. Tissue- and isoform-specific kinetic behavior of the Na,K-ATPase. J Biol Chem 1994;269:16668–16676
- Sahin-Erdemli I, Medford RM, Songu-Mize E. Regulation of Na+,K(+)-ATPase & subunit isoforms in rat tissues during hypertension. Eur J Pharmacol 1995;292:163–171
- Riedl U, Barocka A, Kolem H, et al. Duration of lithium treatment and brain lithium concentration in patients with unipolar and schizoaffective disorder: a study with magnetic resonance spectroscopy. Biol Psychiatry 1997;41:844–850
- Kabakov AY, Karkanias NB, Lenox RH, et al. Synapse specific accumulation of lithium in intracellular microdomains: a model for uncoupling coincidence detection in the brain. Synapse. In press
- Goodnick PJ, Gershon ES. Lithium. In: Lajtha A, ed. Handbook of Neurochemistry. New York, NY: Plenum Press 1985:103–149
- Bunney WE, Garland-Bunney BL. Mechanism of action of lithium in affective illness: basic and clinical implications. In: Meltzer HY, ed. Psychopharmacology: The Third Generation of Progress. New York, NY: Raven Press; 1987:553–565
- Bunney WE Jr. Neuronal receptor function in psychiatry: strategy and theory. In: Usdin E, Bunney WE Jr, Davis JM, eds. Neuroreceptors: Basic and Clinical Aspects. New York, NY: John Wiley & Sons; 1981:241–255
- Gallager DW, Pert A, Bunney WE Jr. Haloperidol-induced presynaptic dopamine supersensitivity is blocked by chronic lithium. Nature 1978;273: 309–312
- Rosenblatt JE, Pert A, Layton B, et al. Chronic lithium reduced ³H-spiroperidol binding in rat striatum. Eur J Pharmacol 1980;67:321–322
- Verimer T, Goodale DB, Long JP, et al. Lithium effects on haloperidolinduced pre- and postsynaptic dopamine receptor supersensitivity. J Pharm Pharmacol 1980;32:665–666
- Bloom FE, Baetge G, Deyo S, et al. Chemical and physiological aspects of the actions of lithium and antidepressant drugs. Neuropharmacology 1983; 22:359–365
- Klawans HL, Weiner WJ, Nausieda PA. The effect of lithium on an animal model of tardive dyskinesia. Prog Neuropsychopharmacol 1976;1:53–60
- 36. Staunton DA, Magistretti PJ, Shoemaker WJ, et al. Effects of chronic lithium treatment on dopamine receptors in the rat corpus striatum, I: locomotor activity and behavioral supersensitivity. Brain Res 1982;232:391–400
- 37. Staunton DA, Magistretti PJ, Shoemaker WJ, et al. Effects of chronic lithi-

um treatment on dopamine receptors in the rat corpus striatum, II: no effect on denervation or neuroleptic-induced supersensitivity. Brain Res 1982; 232:401–412

- Allikmets LH, Stanley M, Gershon S. The effect of lithium on chronic haloperidol enhanced apomorphine aggression in rats. Life Sci 1979;25: 165–170
- Huey LY, Janowshy DS, Judd LL, et al. Effects of lithium carbonate on methylphenidate-induced mood, behavior, and cognitive processes. Psychopharmacology (Berl) 1981;73:161–164
- Pert A, Rosenblatt JE, Sivit C, et al. Long-term treatment with lithium prevents the development of dopamine receptor supersensitivity. Science 1978;201:171–173
- van Kammen DP, Docherty JP, Marder SR, et al. Lithium attenuates the activation-euphoria but not the psychosis induced by d-amphetamine in schizophrenia. Psychopharmacology (Berl) 1985;87:111–115
- Friedman E, Wang H-Y. Effect of chronic lithium treatment on 5-hyroxytryptamine autoreceptors and release of 5-[3H]hydroxytryptamine from rat brain cortical, hippocampal, and hypothalamic slices. J Neurochem 1988;50:195–201
- Goodwin GM, DeSouza RJ, Wood AJ, et al. Lithium decreases 5-HT_{1A} and 5-HT₂ receptor and α₂ adrenoceptor mediated function in mice. Psychopharmacology (Berl) 1986;90:482–487
- Hotta I, Yamawaki S. Possible involvement of presynaptic 5-HT autoreceptors in effect of lithium on 5-HT release in hippocampus of rat. Neuropharmacology 1988;27:987–992
- 45. Mork A, Geisler A. Effects of GTP on hormone-stimulated adenylate cyclase activity in cerebral cortex, striatum, and hippocampus from rats treated chronically with lithium. Biol Psychiatry 1989;26:279–288
- Newman ME, Drummer D, Lerer B. Single and combined effects of desimipramine and lithium on serotonergic receptor number and second messenger function in rat brain. J Pharmacol Exp Ther 1990;252:826–831
- Wang HY, Friedman E. Chronic lithium: desensitization of autoreceptors mediating serotonin release. Psychopharmacology (Berl) 1988;94:312–314
- Blier P, de Montigny C. Short-term lithium administration enhances serotonergic neurotransmission: electrophysiological evidence in the rat CNS. Psychopharmacology (Berl) 1985;113:69–77
- 49. Blier P, de Montigny C, Tardif D. Short-term lithium treatment enhances responsiveness of postsynaptic 5-HT_{1A} receptors without altering 5-HT autoreceptor sensitivity: an electrophysiological study in the rat brain. Synapse 1987;1:225–232
- 50. Dilsaver SC, Coffman JA. Cholinergic hypothesis of depression: a reappraisal. J Clin Psychopharmacol 1989;9:173–179
- Kofman O, Belmaker RH, Grisaru N, et al. *Myo*-inositol attenuates two specific behavioral effects of acute lithium in rats. Psychopharmacol Bull 1991;27:185–190
- Tricklebank MD, Singh L, Jackson A, et al. Evidence that a proconvulsant action of lithium is mediated by inhibition of *myo*-inositol phosphatase in mouse brain. Brain Res 1991;558:145–148
- Evans MS, Zorumski CF, Clifford DB. Lithium enhances neuronal muscarinic excitation by presynaptic facilitation. Neuroscience 1990;38:457–468
- Emerich HM, Zerssen DV, Kissling W, et al. Effect of sodium valproate on mania. Archives of Psychiatry 1980;229:1–16
- Lloyd KG, Morselli PL, Bartholini G. GABA and affective disorders. Medical Biology 1987;65:159–165
- 56. Squires RF, Saederup E. A review of evidence for GABAergic predominance/glutamatergic deficit as a common etiological factor in both schizophrenia and affective psychoses: more support for a continuum hypothesis of functional psychosis. Neurochem Res 1991;16:1099–1111
- Berrettini WH, Nurnberger JI Jr, Hare T, et al. Plasma and CSF GABA in affective illness. Br J Psychiatry 1982;141:483–487
- Petty F, Kramer GL, Rush AJ. Low plasma γ-aminobutyric acid levels in male patients with depression. Biol Psychiatry 1992;32:354–363
- Post RM, Ballenger JC, Hare TA, et al. Cerebrospinal fluid GABA in normals and patients with affective disorders. Brain Res Bull 1980;5(suppl 2):755–759
- Petty F, Kramer GL, Fulton M, et al. Low plasma GABA is a trait-like marker for bipolar illness. Neuropsychopharmacology 1993;9:125–132
- Berrettini WH, Nurnberger JI Jr, Hare T, et al. Reduced plasma and CSF γ-aminobutyric acid in affective illness: effect of lithium carbonate. Biol Psychiatry 1983;18:185–194
- Berrettini WH, Nurnberger JJ Jr, Hare T, et al. CSF GABA in euthymic manic-depressive patients and controls. Biol Psychiatry 1986;21:842–844
- 63. Petty F, Rush AJ, Davis JM, et al. Plasma GABA predicts acute response to

divalproex in mania. Biol Psychiatry 1996;39:278-284

- Ahluwalia P, Singhal RL. Monoamine uptake into synaptosomes from various regions of rat brain following lithium administration and withdrawal. Neuropharmacology 1981;20:483–487
- Gottesfeld Z, Ebstein BS, Samuel D. Effect of lithium on concentrations of glutamate and GABA levels in amygdala and hypothalamus of rat. Nature 1971;234:124–125
- Maggi A, Enna SJ. Regional alterations in rat brain neurotransmitter systems following chronic lithium treatment. J Neurochem 1980;34:888–892
- 67. Bernasconi R. The GABA hypothesis of affective illness: influence of clinically effective antimanic drugs on GABA turnover. In: Emrich HM, Adenhoff JB, Lux HM, eds. Basic Mechanisms in the Action of Lithium. Amsterdam, The Netherlands: Excerpta Medica; 1982:183–192
- Weiss S, Kemp DE, Bauce L, et al. Kainate receptors coupled to the evoked release of [³H]-γ-aminobutyric acid from striatal neurons in primary culture potentiated by lithium ions. Mol Pharmacol 1990;38:229–236
- 69. Dixon JF, Los GV, Hokin LE. Lithium stimulates glutamate "release" and inositol 1,4,5-triphosphate accumulation via activation of the *N*-methyl-Daspartate receptor in monkey and mouse cerebral cortex slices. Proc Natl Acad Sci U S A 1994;91:8358–8362
- Ormandy GC, Jope RS, Snead OC. Anticonvulsant actions of MK-801 on the lithium-pilocarpine model of status epilepticus in rats. Exp Neurol 1989;106:172–180
- Richter-Levin G, Markham H, Segal M. Spontaneous recovery of deficits in spatial memory and cholinergic potentiation of NMDA in CA₁ neurons during chronic lithium treatment. Hippocampus 1992;2:279–286
- Sivam SP, Krause JE, Takeuchi K, et al. Lithium increases rat striatal β- and gamma-preprotachykinin messenger RNAs. J Pharmacol Exp Ther 1989; 248:1297–1301
- Sivam SP, Takeuchi K, Li S, et al. Lithium increases dynorphin A(1-8) and prodynorphin mRNA levels in the basal ganglia of rats. Brain Res 1988; 427:155–163
- Blancquaert JP, Lefebvre RA, Willems JL. Antiaversive properties of opioids in the conditioned taste aversion test in the rat. Pharmacol Biochem Behav 1987;27:437–441
- Lieblich I, Yirmiya R. Naltrexone reverses a long term depressive effect of a toxic lithium injection on saccharin preference. Physiol Behav 1987;39: 547–550
- Mucha RF, Herz A. Motivational properties of kappa and mu opioid receptor agonists studied with place and taste preference conditioning. Psychopharmacology (Berl) 1985;86:274–280
- 77. Shippenberg TS, Herz A. Influence of chronic lithium treatment upon the motivational effects of opioids: alteration in the effects of mu- but not kappa-opioid receptor ligands. J Pharmacol Exp Ther 1991;256:1101–1106
- Shippenberg TS, Millan MJ, Mucha RF, et al. Involvement of β-endorphin and mu-opioid receptors in mediating the aversive effect of lithium in the rat. Eur J Pharmacol 1988;154:135–144
- Berrettini WH, Nurnberger JI, Scheinin M, et al. Cerebrospinal fluid and plasma monamines and their metabolites in euthymic bipolar patients. Biol Psychiatry 1985;20:257–269
- Berrettini WH, Nurnberger JI Jr, Zerbe RL, et al. CSF neuropeptides in euthymic bipolar patients and controls. Br J Psychiatry 1987;150:208–212
- Allison JH, Stewart MA. Reduced brain inositol in lithium treated rats. Nature: New Biology 1971;233:267–268
- Hallcher LM, Sherman WR. The effects of lithium ion and other agents on the activity of myoinositol-1-phosphatase from bovine brain. J Biol Chem 1980;255:10896–10901
- Batty IH, Downes CP. The inhibition of phosphoinositide synthesis and muscarinic-receptor-mediated phospholipase C activity by Li+ as secondary, selective consequences of inositol depletion in 1321N1 cells. Biochem J 1994;297:529–537
- Sherman WR. Lithium and the phosphoinositide signalling system. In: Birch NJ, ed. Lithium and the Cell. London, England: Academic Press; 1991:121–157
- Batty IH, Downes CP. The mechanism of muscarinic receptor-stimulated phosphatidylinositol resynthesis in 1321N1 astrocytoma cells and its inhibition by Li⁺. J Neurochem 1995;65:2279–2289
- Nahorski SR, Ragan CI, Challiss RAJ. Lithium and the phosphoinositide cycle: an example of uncompetitive inhibition and its pharmacological consequences. Trends Pharmacol Sci 1991;12:297–303
- Nahorski SR, Jenkinson S, Challiss RA. Disruption of phosphoinositide signalling by lithium. Biochem Soc Trans 1992;20:430–434
- 88. Berridge MJ, Downes CP, Hanley MR. Neural and developmental actions

of lithium: a unifying hypothesis. Cell 1989;59:411-419

- Berridge MJ, Downes CP, Hanley MR. Lithium amplifies agonistdependent phosphatidylinositol responses in brain and salivary glands. Biochem J 1982;206:587–595
- Kennedy ED, Challiss RAJ, Ragan CI, et al. Reduced inositol polyphosphate accumulation and inositol supply induced by lithium in stimulated cerebral cortex slices. Biochem J 1990;267:781–786
- Varney MA, Godfrey PP, Drummond AH, et al. Chronic lithium treatment inhibits basal and agonist-stimulated responses in rat cerebral cortex and GH₃ pituitary cells. Mol Pharmacol 1992;4:671–678
- Maslanski JA, Leshko L, Busa WB. Lithium-sensitive production of inositol phosphates during amphibian embryonic mesoderm induction. Science 1992;256:243–245
- Moore GJ, Bebchuk JM, Manji HK. Proton MRS in manic depressive illness: monitoring of lithium induced modulation of brain *myo*-inositol. Society of Neuroscience 1997;23:335–336
- Manji HK, Lenox RH. Long-term action of lithium: a role for transcriptional and posttranscriptional factors regulated by protein kinase C. Synapse 1994;16:11–28
- Watson DG, Lenox RH. Chronic lithium-induced down-regulation of MARCKS in immortalized hippocampal cells: potentiation by muscarinic receptor activation. J Neurochem 1996;67:767–777
- Manji HK, Chen G, Hsiao JK, et al. Regulation of signal transduction pathways by mood-stabilizing agents: implications for the delayed onset of therapeutic efficacy. J Clin Psychiatry 1996;57(suppl 13):34–46
- Becchetti A, Whitaker M. Lithium blocks cell cycle transitions in the first cell cycles of sea urchin embryos, an effect rescued by *myo*-inositol. Development 1997;124:1099–1107
- Jope RS, Williams MB. Lithium and brain signal transduction systems. Biochem Pharmacol 1994;47:429–441
- Anderson PH, Geisler A. Lithium inhibition of forskolin-stimulated adenylate cyclase. Neuropsychobiology 1984;12:1–3
- Ebstein RP, Hermoni M, Belmaker RH. The effect of lithium on noradrenaline-induced cyclic AMP accumulation in rat brain: inhibition after chronic treatment and absence of supersensitivity. J Pharmacol Exp Ther 1980;213:161–167
- 101. Forn J, Valdecasas FG. Effects of lithium on brain adenyl cyclase activity. Biochem Pharmacol 1971;20:2773–2779
- 102. Geisler A, Klysner R, Andersen PH. Influence of lithium in vitro and in vivo on the catecholamine-sensitive cerebral adenylate cyclase systems. Acta Pharmacologica et Toxicologica (Copenh) 1985;56:80–97
- Mork A, Geisler A. Mode of action of lithium on the catalytic unit of adenylate cyclase from rat brain. Pharmacol Toxicol 1987;60:241–248
- Mork A, Geisler A. The effects of lithium in vitro and ex vivo on adenylate cyclase in brain are exerted by distinct mechanisms. Neuropharmacology 1989;28:307–311
- Mork A, Geisler A. Effects of lithium ex vivo on the GTP-mediated inhibition of calcium-stimulated adenylate cyclase activity in rat brain. Eur J Pharmacol 1989;168:347–354
- Mork A, Geisler A. Effects of lithium treatment on agonist-enhanced extracellular concentrations of cyclic AMP in the dorsal hippocampus of freely moving rats. J Neurochem 1995;65:134–139
- 107. Newman ME, Belmaker RH. Effects of hithium in vitro and ex vivo on components of the adenylate cyclase system in membranes from the cerebral cortex of the rat. Neuropharmacology 1987;26:211–217
- Dousa TP. Interaction of lithium with vasopressin-sensitive cyclic AMP system of human renal medulla. Endocrinology 1974;95:1359–1366
- Goldberg H, Clayman P, Skorecki K. Mechanism of Li inhibition on vasopressin-sensitive adenylate cyclase in cultured renal epithelial cells. Am J Physiol 1988;24:F995–F1002
- Tseng FY, Pasquali D, Field JB. Effects of lithium on stimulated metabolic parameters in dog thyroid slices. Acta Endocrinology (Copenhagen) 1989;121:615–620
- Urabe M, Hershman JM, Pang XP, et al. Effect of lithium on function and growth of thyroid cells in vitro. Endocrinology 1991;129:807–814
- 112. Newman M, Klein E, Birmaher B, et al. Lithium at therapeutic concentrations inhibits human brain noradrenaline sensitive cyclic AMP accumulation. Brain Res 1983;278:380–381
- 113. Masana MI, Bitran JA, Hsiao JK, et al. Lithium effects on noradrenergic linked adenylate cyclase activity in intact rat brain: an in vivo microdialysis study. Brain Res 1991;538:333–336
- 114. Godfrey PP, McClue SJ, White AM, et al. Subacute and chronic in vivo lithium treatment inhibits agonist- and sodium fluoride-stimulated inosi-

tol phosphate production in rat cortex. J Neurochem 1989;52:498-506

- Song L, Jope R. Chronic lithium treatment impairs phosphatidylinositol hydrolysis in membranes from rat brain regions. J Neurochem 1992;58: 2200–2206
- 116. Avissar S, Schreiber G, Danon A, et al. Lithium inhibits adrenergic and cholinergic increases in GTP binding in rat cortex. Nature 1988;331: 440–442
- Ellis J, Lenox RH. Receptor coupling to G proteins: interactions not affected by lithium. Lithium 1991;2:141–147
- Manji HK, Chen G, Shimon H, et al. Guanine nucleotide-binding proteins in bipolar affective disorder: effects of long-term lithium treatment. Arch Gen Psychiatry 1995;52:135–144
- Manji HK. G proteins: implications for psychiatry. Am J Psychiatry 1992; 149:746-760
- Masana MI, Bitran JA, Hsiao JK, et al. In vivo evidence that lithium inactivates G_i modulation of adenylate cyclase in brain. J Neurochem 1992; 59:200–205
- 121. Bebchuk JM, Arfken CL, Dolan-Manji S, et al. A preliminary investigation of protein kinase C inhibitor (tamoxifen) in the treatment of acute mania. Presented at the American College of Neuropsychopharmacology; December 8–12, 1997; Waikoloa, Hawaii
- Greenwood AF, Jope RS. Brain G-protein proteolysis by calpain: enhancement by lithium. Brain Res 1994;636:320–326
- Li PP, Tam YK, Young LT, et al. Lithium decreases G_s, G_{i1} and G_{i2} & subunit mRNA levels in rat cortex. Eur J Pharmacol 1991;206:165–166
- Hsiao JK, Colison J, Bartko JJ, et al. Monoamine neurotransmitter interactions in drug-free and neuroleptic-treated schizophrenics. Arch Gen Psychiatry 1993;50:606–614
- 125. Colin SF, Chang HC, Mollner S, et al. Chronic lithium regulates the expression of adenylate cyclase and G_i-protein αsubunit in rat cerebral cortex. Proc Natl Acad Sci U S A 1991;88:10634–10637
- Thiele EA, Eipper BA. Effect of secretagogues on components of the secretory system in AtT-20 cells. Endocrinology 1990;126:809–817
- Young LT, Woods CM. Mood stabilizers have differential effects on endogenous ADP ribosylation in C6 glioma cells. Eur J Pharmacol 1996; 309:215–218
- Nestler EJ, Terwilliger RZ, Duman RS. Regulation of endogenous ADPribosylation by acute and chronic lithium in rat brain. J Neurochem 1995; 64:2319–2324
- 129. Huang KP. The mechanism of protein kinase C activation. Trends Neurosci 1989;12:425–432
- Nishizuka Y. Intracellular signaling by hydrolysis of phospholipids and activation of protein kinase C. Science 1992;258:607–614
- 131. Stabel S, Parker PJ. Protein kinase C. Pharmacol Ther 1991;51:71–95
- Tanaka C, Nishizuka Y. The protein kinase C family for neuronal signaling. Annu Rev Neurosci 1994;17:551–567
- 133. Manji HK, Etcheberrigaray R, Chen G, et al. Lithium dramatically decreases membrane-associated PKC in the hippocampus: selectivity for the α isozyme. J Neurochem 1993;61:3203–3210
- Manji HK, Bersudsky U, Chen G, et al. Modulation of PKC isozymes and substrates by lithium: the role of *myo*-inositol. Neuropsychopharmacology 1996;15:370–381
- 135. Leli U, Hauser G. Lithium modifies diacylglycerol levels and protein kinase C in neuroblastoma cells. In: Abstracts of the 8th International Conference on Second messengers and phosphoproteins; August 3–6, 1992; Glasgow, Scotland
- Li X, Jope RS. Selective inhibition of the expression of signal transduction proteins by lithium in nerve growth factor-differentiated PC12 cells. J Neurochem 1995;65:2500–2508
- 137. Chen G, Manji HK, Hawver DB, et al. Chronic sodium valproate selectively decreases protein kinase C α and ϵ in vitro. J Neurochem 1994;63: 2361–2364
- Huang FL, Yoshida Y, Cunha-Melo JR, et al. Differential down-regulation of protein kinase C isozymes. J Biol Chem 1989;264:4238–4243
- Isakov N, McMahon P, Altman A. Selective post-transcriptional downregulation of protein kinase C isozymes in Leukemic T cells chronically treated with phorbol ester. J Biol Chem 1990;265:2091–2097
- Oda T, Shearman MS, Nishizuka Y. Synaptosomal protein kinase C subspecies, B: down-regulation promoted by phorbol ester and its effect on evoked norepinephrine release. J Neurochem 1991;56:1263–1269
- 141. Watson DG, Wainer BH, Lenox RH. Phorbol ester- and retinoic acidinduced regulation of the protein kinase C substrate MARCKS in immortalized hippocampal cells. J Neurochem 1994;63:1666–1674

- Lenox RH, Watson DG, Patel J, et al. Chronic lithium administration alters a prominent PKC substrate in rat hippocampus. Brain Res 1992;570: 333–340
- Aderem A. The MARCKS brothers: a family of protein kinase C substrates. Cell 1992;71:713–716
- Blackshear PJ. The MARCKS family of cellular protein kinase C substrates. J Biol Chem 1993;268:1501–1504
- 145. Lenox RH, McNamara RK, Watterson JM, et al. Myristoylated alaninerich C kinase substrate (MARCKS): a molecular target for the therapeutic action of mood stabilizers in the brain? J Clin Psychiatry 1996;57(suppl 13):23–31
- 146. Watson DG, Watterson JM, Lenox RH. Sodium valproate down-regulates the myristoylated alanine-rich C kinase substrate (MARCKS) in immortalized hippocampal cells: a property of PKC-mediated mood stabilizers. J Pharmacol Exp Ther. In press
- 147. Bohmann D. Transcription factor phosphorylation: a link between signal transduction and the regulation of gene expression. Cancer Cells 1990;2: 337–343
- Hunter T, Karin M. The regulation of transcription by phoshphorylation. Cell 1992;70:377–387
- 149. Divish MM, Sheftel G, Boyle A, et al. Differential effect of lithium on fos protooncogene expression mediated by receptor and postreceptor activators of protein kinase C and cyclic adenosine monophosphate: model for its antimanic action. J Neurosci Res 1991;28:40–48
- 150. Gao XM, Fukamauchi F, Chuang DM. Long-term biphasic effects of lithium treatment on phospholipase C-coupled M3-muscarinic acetylcholine receptors in cultured cerebellar granule cells. Neurochem Int 1993;22: 395–403
- 151. Kalasapudi VD, Sheftel G, Divish MM, et al. Lithium augments fos protooncogene expression in PC12 pheochromocytoma cells: implications for therapeutic action of lithium. Brain Res 1990;521:47–54
- Miller JC, Mathe AA. Basal and stimulated c-fos mRNA expression in the rat brain: effect of chronic dietary lithium. Neuropsychopharmacology 1997;16:408–418
- 153. Kislauskis E, Dobner PR. Mutually dependent response elements in the cis-regulatory region of the neurotensin/neuromedin N gene integrate environmental stimuli in PC12 cells. Neuron 1990;4:783–795
- 154. Pfeiffer A, Veilleux S, Barden N. Antidepressant and other centrally acting drugs regulate glucocorticoid receptor messenger RNA levels in rat brain. Psychoneuroendocrinology 1991;16:505–515
- 155. Weiner ED, Kalaaspudi VD, Papolos DF, et al. Lithium augments pilocarpine-induced fos gene expression in brain. Brain Res 1991;553: 117–122
- 156. Zachrisson O, Mathe AA, Stenfors C, et al. Regional-specific effects of chronic lithium on administration on neuropeptide Y and somatostatin mRNA expression in the rat brain. Neurosci Lett 1995;194:89–92
- 157. Chen G, Yuan P, Wawyer DB, et al. Increase in AP-1 transcription factor DNA binding activity by valproic acid. Neuropsychopharmacology 1997; 16:238–245
- Unlap MT, Jope RS. Lithium attenuates nerve growth factor-induced activation of AP-1 DNA binding activity in PC12 cells. Neuropsychopharmacology 1997;17:12–17
- 159. Bond PA, Brooks BA, Judd A. The distribution of lithium, sodium and magnesium in rat brain and plasma after various periods of administration of lithium in the diet. Br J Pharmacol 1975;53:235–239
- Ebadi MS, Simmons VJ, Hendrickson MJ, et al. Pharmakokinetics of lithium and its regional distribution in rat brain. Eur J Pharmacol 1974; 27:324–329
- Edelfors S. Distribution of sodium, potassium and lithium in the brain of lithium-treated rats. Acta Pharmacologica et Toxicologica 1975;37: 387–392
- Heurteaux C, Baumann N, Lachapelle F, et al. Lithium distribution in the brain of normal mice and of "quacking" dysmyelinating mutants. J Neurochem 1986;46:1317–1321
- Mukherjee BP, Bailey PT, Pradhan SN. Temporal and regional differences in brain concentrations of lithium in rats. Psychopharmacology (Berl) 1976;48:119–121
- 164. Nelson SC, Herman MM, Bensch KG, et al. Localization and quantitation of lithium in rat tissue following intraperitoneal injections of lithium chloride, II: Brain. J Pharm Exp Ther 1980;212:11–15
- Ramaprasad S, Newton JEO, Cardwell D, et al. In vivo ⁷Li NMR imaging and localized spectroscopy of rat brain. Magnetic Resonance Medicine 1992;25:308–318

- 166. Sandner G, Di Scala G, Oberling P, et al. Distribution of lithium in the rat brain after a single administration known to elicit aversive effects. Neurosci Lett 1994;166:1-4
- 167. Savalainen KM, Hirvonen MR, Tarhanen J, et al. Changes in cerebral inositol-1-phosphate concentrations in LiCl-treated rats: regional and strain differences. Neurochem Res 1990;15:541-545
- 168. Smith DF, Amdisen A. Lithium distribution in rat brain after longterm central administration by minipump. J Pharm Phamacol 1981;33: 805-806
- 169. Spirtes MA. Lithium levels in monkey and human brain after chronic, therapeutic, oral dosage. Pharmacol Biochem Behav 1976;5:143-147
- 170. Thellier M, Wissocq JC, Heurteaux C. Quantitative microlocation of lithium in the brain by a (n, o) nuclear reaction. Nature 1980;283:299–302
- 171. Allison JH, Boshans RL, Hallcher LM, et al. The effects of lithium on myo-inositol levels in layers of frontal cerebral cortex, in cerebellum, and in corpus callosum of the rat. J Neurochem 1980;34:456-458
- 172. Johnson RD, Minneman KP. q-adrenergic receptors and stimulation of [3H]inositol metabolism in rat brain: regional distribution and parallel inactivation. Brain Res 1985;341:7-15

- 173. Jope RS, Song L, Li PP, et al. The phosphoinositide signal transduction system is impaired in bipolar affective disorder brain. J Neurochem 1996; 66:2402-2409
- 174. Rooney TA, Nahorski SR. Regional characterization of agonist and depolarization-induced phosphoinositide hydrolysis in rat brain. J Pharmacol Exp Ther 1986;239:873-880
- 175. Sherman WR, Gish BG, Honchar MP, et al. Effects of lithium on phosphoinositide metabolism in vivo. Federation Proceedings 1986;45: 2639-2646
- 176. Gani D, Downes CP, Bramham J. Lithium and myo-inositol homeostasis. Biochim Biophys Acta 1993;1177:253-269
- 177. Heacock AM, Seguin EB, Agranoff BW. Measurement of receptoractivated phosphoinositide turnover in rat brain: nonequivalence of inositol phosphate and CDP-diacylglycerol formation. J Neurosci 1993;60: 1087-1092
- 178. Sarri E, Picatoste F, Claro E. Neurotransmitter specific profiles of inositol phosphates in rat brain cortex: relation to the mode of receptor activation of phosphoinositide phospholipase C. J Pharmacol Exp Ther 1995;272:

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