A High-Density Whole-Genome Association Study Reveals That *APOE* Is the Major Susceptibility Gene for Sporadic Late-Onset Alzheimer's Disease

Keith D. Coon, Ph.D.; Amanda J. Myers, Ph.D.; David W. Craig, Ph.D.;
Jennifer A. Webster, B.A.; John V. Pearson, B.Sc.; Diane Hu Lince, Ph.D.; Victoria L. Zismann, M.S.;
Thomas G. Beach, M.D.; Doris Leung, M.D.; Leslie Bryden, M.S.; Rebecca F. Halperin, B.Sc.;
Lauren Marlowe, B.Sc.; Mona Kaleem, B.Sc.; Douglas G. Walker, Ph.D.; Rivka Ravid, Ph.D.;
Christopher B. Heward, Ph.D.; Joseph Rogers, Ph.D.; Andreas Papassotiropoulos, M.D.;
Eric M. Reiman, M.D.; John Hardy, Ph.D.; and Dietrich A. Stephan, Ph.D.

Objective: While the apolipoprotein E (*APOE*) ε4 allele is a well-established risk factor for late-onset Alzheimer's disease (AD), initial genome scans using microsatellite markers in late-onset AD failed to identify this locus on chromosome 19. Recently developed methods for the simultaneous assessment of hundreds of thousands of single nucleotide polymorphisms (SNPs) promise to help more precisely identify loci that contribute to the risk of AD and other common multigenic conditions. We sought here to demonstrate that more precise identification of loci that are associated with complex, multigenic genetic disorders can be achieved using ultra-highdensity whole-genome associations by demonstrating their ability to identify the APOE locus as a major susceptibility gene for late-onset AD, despite the absence of SNPs within the APOE locus itself, as well as to refine odds ratios (ORs) based on gold-standard phenotyping of the study population.

Method: An individualized genome-wide association study using 502,627 SNPs was performed in 1086 histopathologically verified AD cases and controls to determine the OR associated with genes predisposing to Alzheimer's disease.

Results: As predicted, ultra-high-density SNP genotyping, in contrast to traditional microsatellite-based genome screening approaches, precisely identified the *APOE* locus as having a significant association with late-onset AD. SNP rs4420638 on chromosome 19, located 14 kilobase pairs distal to the *APOE* ϵ 4 variant, significantly distinguished between AD cases and controls (Bonferroni corrected p value = 5.30×10^{-34} , OR = 4.01) and was far more strongly associated with the risk of AD than any other SNP of the 502,627 tested.

Conclusion: This study provides empirical support for the suggestion that the APOE locus is the major susceptibility gene for late-onset AD in the human genome, with an OR significantly greater than any other locus in the human genome. It also supports the feasibility of the ultra-high-density whole-genome association approach to the study of AD and other heritable phenotypes. These whole-genome association studies show great promise to identify additional genes that contribute to the risk of AD. (J Clin Psychiatry 2007;68:613–618)

Received Oct. 20, 2006; accepted Jan. 30, 2007. From the Neurogenomics Division, Translational Genomics Research Institute, Phoenix, Ariz. (Drs. Coon, Craig, Lince, Papassotiropoulos, Reiman, and Stephan; Mss. Webster, Zismann, and Halperin; and Mr. Pearson); the Department of Psychiatry and Behavioral Sciences, University of Miami, Miller School of Medicine, Miami, Fla. (Dr. Myers); the Sun Health Research Institute, Sun City, Ariz. (Drs. Beach, Walker, and Rogers); the Laboratory of Neurogenetics, National Institute on Aging, National Institutes of Health, Bethesda, Md. (Drs. Leung and Hardy and Mss. Bryden, Marlowe, and Kaleem); the Royal Dutch Academy of Sciences, Amsterdam, the Netherlands (Dr. Ravid); Kronos Science Laboratories, Phoenix, Ariz. (Dr. Heward); the Division of Psychiatry Research, University of Zurich, Zurich, Switzerland (Dr. Papassotiropoulos); and Banner Alzheimer's Institute, Phoenix (Dr. Reiman); the Department of Psychiatry, University of Arizona, Tucson (Dr. Reiman); and Arizona Alzheimer's Consortium, Phoenix (Drs. Reiman and Stephan), Ariz.

Support statements and acknowledgments appear at the end of this article.

Corresponding author and reprints: Dietrich A. Stephan, Ph.D., Director, Neurogenomics Division, TGen, Translational Genomics Research Institute, 400 N. Fifth St., Suite 1600, Phoenix, AZ 85004 (e-mail: dstephan@tgen.org).

lzheimer's disease (AD) is a devastating neuro-degenerative disorder typically characterized by progressively disabling impairments in memory and other cognitive domains but also by noncognitive behavioral symptoms. AD preferentially affects individuals over 60 years, with prevalence rates as high as 40% in nonagenarians. Sporadic AD is multifactorial and genetically complex. Twin studies suggest that genetic factors may account for as much as 80% of the disease risk. While several genes and genetic polymorphisms therein have been suggested as AD susceptibility factors, the only well-verified susceptibility gene for AD is the apolipoprotein E (*APOE*) ε4 allele. 3.4

Located on chromosome 19, the *APOE* gene encodes the apolipoprotein E protein, which is known to play a central role in the regulation of the cholesterol and triglyceride metabolism.⁵ The 3 common *APOE* alleles ($\varepsilon 2$, $\varepsilon 3$, and $\varepsilon 4$) are distinguished from each other on the basis of 2 single nucleotide polymorphisms (SNPs), resulting in 2 amino acid changes at positions 112 and 158. The *APOE* $\varepsilon 3$ allele is characterized by cysteine at positions 112 and

158, the APOE $\varepsilon 4$ allele by cysteine at position 112 and arginine at position 158, and the APOE ε4 allele by arginine at both positions. A significant association of the APOE £4 allele with AD was initially demonstrated in 1993.³ This finding was replicated in numerous studies and is now considered the best-established genetic association with AD.6 The magnitude of the effect of the APOE ε4 allele as a risk factor for AD is influenced by a person's age and ethnicity. The highest odds ratios (ORs) are detected in the Japanese population (OR = 5.6 for APOE ε 4 allele heterozygous, OR = 33.1 for APOE ε 4 allele homozygous). In Caucasian populations through meta-analysis of multiple studies (with primarily antemortem study populations, which have, at best, 90% diagnostic accuracy), APOE & 4-heterozygous individuals have been calculated to have an approximately 3-fold increased risk and homozygous persons to have an approximately 15-fold increased risk for developing AD by age 75 years compared with APOE ε3-homozygous individuals. ⁴ Although the association between the APOE ε4 allele and AD is consistent and strong, initial genome scans using microsatellite markers in late-onset AD failed to identify this locus on chromosome 19.7

The completion of the human genome sequencing effort, the cataloging of the majority of interindividual DNA variants (SNPs) in public databases, and the evolution of massively parallelized SNP genotyping technologies has permitted the undertaking of non-hypothesisdriven whole-genome association studies. Alleles of every gene in the genome can now be queried for imbalances between cases and controls. It has been proposed that at least 300,000 SNPs across the genome are required to perform a genome-wide association study in an outbred population,⁸ and technologies have only recently become available to permit this kind of study. One such technology, the 500K GeneChip from Affymetrix (Affymetrix, Santa Clara, Calif.), was used in the present study. According to the manufacturer, the 500K array allows ~80% of the genome in the Caucasian population to be queried for association signals. Interestingly, the APOE functional variants are not represented on the GeneChip. However, if our whole-genome approach has adequately high resolution (i.e., SNPs close enough to the actual SNP involved in the susceptibility to the phenotype; high linkage disequilibrium [LD]) and adequate statistical power, the APOE locus should be ascertained with SNPs on the GeneChip other than, but close to, the APOE ε2/3/4 variant.

We have investigated the hypothesis that ultra-highdensity SNP genotyping approaches in histopathologically confirmed and clinically assessed samples will identify loci that have been largely undetectable using traditional microsatellite-based protocols. To test the feasibility of this approach, we sought to demonstrate the ability of our high-density whole-genomic association method to distinguish AD cases from controls as well as permit us to confirm the widespread, but not yet empirically confirmed, belief that the *APOE* locus is more strongly associated with the risk of AD than any other locus in the human genome. Finally, we have been able to refine the OR of the *APOE* $\varepsilon 2/3/4$ carriers relative to AD.

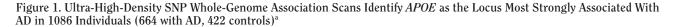
METHOD

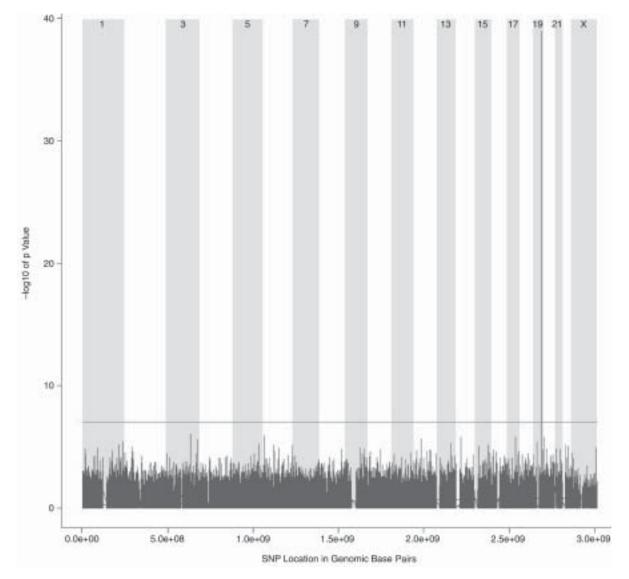
Samples

DNA samples were extracted from brain tissue of 1086 brain donors who were at least 65 years of age at the time of their death. The donors included 664 patients who satisfied clinical and neuropathologic criteria for the diagnosis of AD and 422 persons with no known AD symptomatology or pathology (controls). Their brain tissue and neuropathologic diagnoses were supplied by investigators from 12 National Institute on Aging Alzheimer's Disease Centers in accordance with an agreement with these Centers, the National Institute on Aging, and the National Alzheimer's Coordinating Center. Additional postmortem samples were received from Sun Health Research Institute and the Netherlands Brain Bank. Eighteen percent of the patients and 16% of controls were assessed using Consortium to Establish a Registry for Alzheimer's Disease (CERAD) criteria, which provide information about neuritic plaque density; 59% of the patients and 75% of the controls were also assessed using Braak and Braak staging, 10 which assesses the distribution of neurofibrillary tangles. APOE genotypes were obtained by either pyrosequencing¹¹ or restriction fragment length polymorphism analysis.¹² The patients included 362 females and 302 males with a mean \pm SD age of 82 \pm 7.7 years at the time of death. The controls included 255 females and 167 males with a mean \pm SD age of 79 \pm 11.0 years at the time of death. No population stratification was present between cases vs. controls using the genomic control method with 50 randomly chosen SNPs.¹³

Affymetrix 500K GeneChip SNP Genotyping and Analysis

Array-based SNP genotyping. Each of the 1086 brain tissue samples was processed using the Gentra DNA isolation protocol (QIAGEN Inc., Valencia, Calif.) to obtain high molecular weight DNA, which was then diluted to 50 ng/mL with reduced Tris-ethylenediaminetetraacetate (Tris/EDTA; TE) buffer. Samples were processed as described in the Mapping 500K Protocol (Affymetrix). Briefly, quality and relative concentration of each pool or sample was assessed on 2% Tris-acetate-EDTA (TAE) agarose gel; 250 ng (5 mL) of DNA from each sample was digested in parallel with 10 units of Nsp I and Sty I restriction enzymes (New England Biolabs, Beverly, Mass.) for 2 hours at 37°C. Enzyme specific adaptor oligonucle-otides were then ligated onto the digested ends with T4





^aThe 500K GeneChip unambiguously identified the *APOE* locus through a non–hypothesis-driven scan. The identified SNP rs4420638 is ~14 kb distal to the *APOE* locus and was found to have, by far, the most significant association p value via Fisher exact test. Abbreviations: AD = Alzheimer's disease, SNP = single nucleotide polymorphism.

DNA Ligase (Invitrogen, Carlsbad, Calif.) for 3 hours at 16°C. After dilution with water, 10 mL of the diluted ligation reactions were subjected to polymerase chain reaction (PCR). PCR was performed using Titanium Taq DNA Polymerase (BD Biosciences, San Jose, Calif.) in the presence of 100 mM PCR primer 002 (Affymetrix), 350 mM each dNTP, 1M G-C melt (Clontech, Mountain View, Calif.), and 1X Titanium Taq PCR Buffer (BD Biosciences). Cycling parameters were as follows: initial denaturation at 94°C for 3 minutes, denaturation at 94°C for 30 seconds, annealing at 60°C for 30 seconds, and extension at 68°C for 15 seconds repeated a total of 30 times,

with a final extension at 68°C for 7 minutes. PCR products from 3 reactions were combined and purified using the Clontech Clean-Up Kit 96-well PCR purification plates (Clontech) according to the manufacturer's directions. PCR products were then verified to migrate at an average size between 200–800 base pairs (bps) using 2% TAE gel electrophoresis. Ninety micrograms of purified PCR products were then fragmented using 0.25 units of DNAase I (Invitrogen) at 37°C for 35 minutes. Complete fragmentation of the products to an average size less than 180 bps was verified using 2% TAE gel electrophoresis. Following fragmentation, the DNA was end

labeled using 105 units of terminal deoxynucleotidyl transferase (Invitrogen) at 37°C for 4 hours. The labeled DNA was then hybridized onto the respective Mapping 500K array at 49°C for 18 hours at 60 rpm. The hybridized array was washed, stained, and scanned according to the manufacturer's (Affymetrix) instructions. Genotypes were extracted using the SNiPer-HD¹⁴ software. All data are stored in a GeneChip Operating Software (Affymetrix) enterprise server at Translational Genomics Research Institute, Phoenix, Ariz. Raw data for the 300 kilobase pairs (kb) encompassing the *APOE* locus are available at http://www.tgen.org/neurogenomics/data.

Statistical analysis. Perl scripts were constructed to intercalate, corresponding to chromosomal position, the 502,627 SNPs that were collectively genotyped on the Sty I and Nsp I arrays for each sample (http://bioinformatics.tgen.org/software/tgen-array/). Calculation of an SNP's allelic frequency distribution between cases and controls was based on the Fisher exact test using each SNP sequentially. Bonferroni correction was performed on the resultant p values, and all SNPs were plotted across the genome using R. The APOE locus had the highest corrected significance level and thus the strongest single allelic association with AD.

Haplotyping and Linkage Disequilibrium Mapping

LD mapping of the 300 kb surrounding the *APOE* locus was performed importing genotypes into the Haploview program v. 3.3 (Broad Institute, Cambridge, Mass.). When variants of 2 genetic loci are in strong LD (measured by D' statistic), the variant seen at one locus is predictive of the variant found at the other on an individual chromosome. LD thus represents the likelihood that 2 genetic markers will be inherited together. Pairwise LD values (as measured by D') for each pair of SNPs across the 300 kb interval were calculated using the Haploview software and plotted (see Figure 2).¹⁵

RESULTS

Unlike traditional microsatellite-based genome screening approaches, the ultra-high-density SNP genotyping performed herein was able to precisely identify the *APOE* locus as having a significant association with late-onset AD. Whole-genome scans of 502,627 SNPs were performed for each of the 1086 samples. The most significant SNP identified, rs4420638 (uncorrected p value = 1.06×10^{-39} , Bonferonni corrected p value for multiple hypothesis testing = 5.30×10^{-34} based on 500,000 comparisons), was located on chromosome 19, 14 kb from the *APOE* ϵ 4 variant. Figure 1 displays the magnitude of this association peak in comparison to other SNP p values across the entire human genome. The specific *APOE* ϵ 4 functional variant is not included on the 500K GeneChip but has been repeatedly identified as a promi-

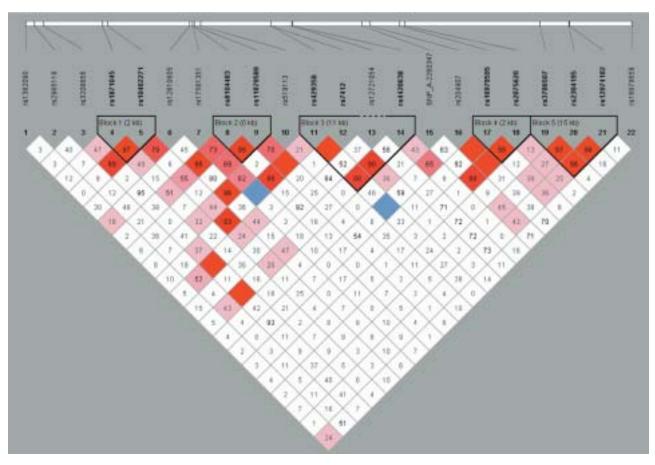
nent genetic risk factor in AD.¹⁷ Figure 2 illustrates the LD (as measured by D') between rs4420638 and APOE locus as defined by the 2 functional variants that define $\varepsilon 2/3/4$ status. Haplotype block 3 (Figure 2) is composed of rs4420638 and the 2 APOE SNPs: rs429358 and rs7412 at a D' = 0.86 and 0.90, respectively (rs12721054 being uninformative with a minor allele frequency of 0.0060). Pairwise D' values for all SNPs in the surrounding ~300 kb are illustrated. The APOE locus would have been missed with the 500K array if not for a single SNP in LD with the functional variant, and speaks to the need for increased SNP density for complete coverage. This locus was 27 orders of magnitude more significant than the next most significant SNP by p value. Table 1 depicts refined odds ratios in a large sampling of very carefully phenotyped study samples, which illustrate the increase in effect of APOE genotype in AD predisposition. As an example, previous meta-analyses have indicated an OR for APOE ε4 homozygotes of 14.9⁴ and 15.6, ¹⁸ whereas here we calculate the OR as 25.3 through only including both histopathologically verified cases and controls who were also assessed for dementia prior to death.

DISCUSSION

The *APOE* gene, known to mediate the regulation of cholesterol and triglyceride metabolism, has been repeatedly implicated in the pathogenesis of AD. $^{3.4,6.16}$ The *APOE* gene confers differential susceptibility to AD etiology depending on the combination of the 3 alleles ($\epsilon 2$, $\epsilon 3$, and $\epsilon 4$) as well as the age and ethnicity of the person. The $\epsilon 4$ allele is most highly associated with AD at a large range of ages and in all ethnic groups. Thus, despite the inability of microsatellite-based genome screens to identify a locus on chromosome 19, the *APOE* gene is consistently and strongly associated with late-onset AD and is considered the single most important genetic factor in AD.

Three major conclusions can be reached based on the results of this study. First, whole-genome association studies using carefully phenotyped cases and controls, when rigorously controlled, adequately powered, and performed using a sufficiently reliable and dense set of markers, can indeed identify the allelic variants that contribute risk toward complex disease. Second, the rare allele of rs4420638, the only informative SNP on the 500K array in LD with the APOE locus, is strongly associated with sporadic late-onset AD, having a p value of 1.06×10^{-39} and an OR of 4.01, the highest (in both cases) across the entire human genome in this sampling. Using rs4420638 as an example, however, caution is warranted about making claims regarding holistic extraction of association findings, as this is the only SNP that detected the major known APOE association and speaks to the need for increased density on SNP arrays even for less-ancient outbred populations. Third, we have refined the ORs associ-

Figure 2. Disequilibrium Values for Each Pair of Single Nucleotide Polymorphisms Across the ~ 300 Kilobase Pairs Interval Surrounding the APOE Locus



^aAs measured by D'.

^bThe SNP identified in this investigation (rs4420638) was found be in significant linkage disequilibrium with SNPs that characterize the APOE ε2/ε3/ε4 locus, rs429358 and rs7412, with D' values of 0.86 and 0.90, respectively. This figure illustrates this significant linkage disequilibrium between rs4420638 and APOE ε2/ε3/ε4 as well as the density and distribution of assayed SNPs within the 300 kb region surrounding the APOE locus.

Abbreviation and figure legend: LOD = log of the odds. The color code on the plot follows the standard color scheme for Haploview: white boxes, |D'| < 1, LOD < 2; shades of pink/red, |D'| < 1, LOD ≥ 2 ; blue, |D'| = 1, LOD < 2; bright red, |D'| = 1, LOD ≥ 2 .

ated with *APOE* genotype status based on a postmortem sampling, which will ultimately provide more accurate risk assessment when prospectively implemented.

CONCLUSION

The findings from this whole-genome association study, utilizing > 500,000 SNPs in a sample of 1086 histopathologically well-characterized AD cases and age-matched unaffected controls, immediately implicate APOE as the single major pathogenic locus (at this density of genome coverage and using Caucasian-specific population LD and SNP informativeness characteristics) and validate the strategy of whole-genome association studies in complex heritable traits. Ultra-high-density SNP genotyping can be successfully utilized to identify

risk loci in complex diseases when rigorously controlled, adequately powered, and performed using a sufficiently reliable and dense set of markers. In the case of AD, the most prominent risk locus was determined to be *APOE*, which has been repeatedly identified as a major risk factor in AD pathogenesis. This study is the first phase in a larger effort to determine other alleles that are significantly associated with AD.

Support and acknowledgements: Sponsors of this study were Kronos Sciences Laboratories, Phoenix, Ariz.; the Arizona Alzheimer's Disease Core Center, Phoenix, Ariz. (P30 AG19610); the National Alzheimer's Coordinating Center, Seattle, Wash. (U01 AG016976); and the State of Arizona. This work was partially supported by the National Institute on Aging (NIA)/National Institutes of Health Intramural Research Program, Bethesda, Md., U.S.A. Dr. Myers is supported in part by the Johnnie B. Byrd, Sr., Alzheimer's Disease and Research Institute, Tampa, Fla. Dr.

APOE	E			Previously Reported	
Genotype	Controls, N	Cases, N	Odds Ratio (vs. ε3/ε3)	Farrer et al.4	Bertram et al. 18
ε2/ε2	15	4	0.42	0.6	
ε2/ε3	35	13	0.41	0.6	0.6
ε2/ε4	5	17	3.49	2.6	
ε3/ε3	259	214	1.00	1.0	1.0
ε3/ε4	76	287	4.32	3.2	4.3
ε4/ε4	7	113	25.31	14.9	15.6

Papassotiropoulos is supported by the Swiss National Science Foundation, Bern, Switzerland (PP00B-68859). Both Drs. Myers and Hardy would like to thank the Verum Foundation, Munich, Germany. Dr. Craig is supported by the Bisgrove charitable donation, Phoenix, Ariz., and Dr. Stephan is supported by the NIH Neuroscience Blueprint, Bethesda, Md. (U24NS051872) and the State of Arizona. The authors report no other financial disclosure or other affiliations relevant to the subject of this article.

We thank Drs. Creighton Phelps, Ph.D.; Marcelle Morrison-Bogorad, Ph.D.; and Marilyn Miller, Ph.D., from the National Institute on Aging, Bethesda, Md.; and Dr. Walter Kukull, Ph.D., from the National Alzheimer's Coordinating Center, Seattle, Wash., for their assistance in the acquisition of tissue samples and data. None of this work would be possible without the generous participation of the patients and controls and their families. Many data and biomaterials were collected from several NIA-National Alzheimer's Coordinating Centers-funded sites. Marcelle Morrison-Bogorad, Ph.D.; Creighton Phelps, Ph.D.; and Walter Kukull, Ph.D., are thanked for helping to coordinate this collection. The directors, pathologists, and technicians involved include National Institute on Aging: Ruth Seemann, B.Sc., Dan Brady, M.D.; Johns Hopkins Alzheimer's Disease Research Center (NIA grant AG 05146): Juan C. Troncoso, M.D., Olga Pletnikova, M.D.; University of California, Los Angeles (NIA grant P50 AG16570): Harry Vinters, M.D., Justine Pomakian, B.Sc.; The Kathleen Price Bryan Brain Bank, Duke University Medical Center (NIA grant AG05128, National Institute of Neurological Disorders and Stroke grant NS39764, National Institute of Mental Health grant MH60451 also funded by GlaxoSmithKline): Christine Hulette, M.D., Director; Stanford University: Dikran Horoupian, M.D., Ahmad Salehi, M.D., Ph.D.; New York Brain Bank, Taub Institute, Columbia University (NYBB): Jean Paul Vonsattel, M.D.; Massachusetts General Hospital: E. Tessa Hedley-Whyte, M.D., Karlotta Fitch, B.Sc.; University of Michigan (NIH grant P50-AG08671): Roger Albin, M.D., Lisa Bain, B.Sc., Eszter Gombosi, B.Sc.; University of Kentucky: William Markesbery, M.D., Sonya Anderson, B.Sc.; Mayo Clinic, Jacksonville: Dennis W. Dickson, M.D., Natalie Thomas, B.Sc.; University of Southern California: Caroll A. Miller, M.D., Jenny Tang, M.S., Dimitri Diaz, B.Sc.; Washington University, St. Louis Alzheimer's Disease Research Center: Dan McKeel, M.D., John C. Morris, M.D., Eugene Johnson, Jr., Ph.D., Virginia Buckles, Ph.D., Deborah Carter, B.Sc.; University of Washington, Seattle: Thomas Montine, M.D., Ph.D., Aimee Schantz, M.Ed.; University of Pennsylvania School of Medicine, Alzheimer's Disease Research Center (NIH grant): John Q. Trojanowski, M.D., Virginia M. Lee, M.D., Vivianna Van Deerlin, M.D., Terry Schuck, B.Sc.; Boston University Alzheimer's Disease Research Center (NIH grant P30-AG13846): Ann C. McKee, B.Sc., Carol Kubilus, B.Sc.; Sun Health Research Institute, Arizona: Joseph Rogers, Ph.D., Thomas G. Beach, M.D., Ph.D., Lucia I. Sue, B.Sc.; Emory University: Bruce H. Wainer, M.D., Ph.D., Marla Gearing, Ph.D.; University of Texas, Southwestern Medical School: Charles L. White III, M.D., Roger Rosenberg, B.Sc., Marilyn Howell, B.Sc., Joan Reisch, B.Sc.; University of California, Davis: William Ellis, M.D., Mary Ann Jarvis, B.Sc.; Rush University Medical Center, Rush Alzheimer's Disease Center: David A. Bennett, M.D., Julie A. Schneider, M.D., M.S., Karen Skish, M.S., PA(ASCP)MT, Wayne T. Longman, B.Sc.; University of Miami/NPF Brain Endowment Bank: Deborah C. Mash, M.D., Margaret J. Basile, B.Sc., Mitsuko Tanaka, B.Sc. All relevant disclosures are noted for acknowledged persons.

REFERENCES

- Evans DA, Funkenstein HH, Albert MS, et al. Prevalence of Alzheimer's disease in a community population of older persons: higher than previously reported. JAMA 1989;262:2551–2556
- Gatz M, Reynolds CA, Fratiglioni L, et al. Role of genes and environments for explaining Alzheimer disease. Arch Gen Psychiatry 2006;63: 168–174
- Saunders AM, Strittmatter WJ, Schmechel D, et al. Association of apolipoprotein E allele epsilon 4 with late-onset familial and sporadic Alzheimer's disease. Neurology 1993;43:1467–1472
- Farrer LA, Cupples LA, Haines JL, et al. Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease: a meta-analysis. APOE and Alzheimer Disease Meta Analysis Consortium. JAMA 1997;278:1349–1356
- Breslow JL, Zannis VI, SanGiacomo TR, et al. Studies of familial type III hyperlipoproteinemia using as a genetic marker the apoE phenotype E2/2. J Lipid Res 1982;23:1224–1235
- Papassotiropoulos A, Fountoulakis M, Dunckley T, et al. Genetics, transcriptomics, and proteomics of Alzheimer's disease. J Clin Psychiatry 2006;67:652–670
- Kehoe P, Wavrant-De Vrieze F, Crook R, et al. A full genome scan for late onset Alzheimer's disease. Hum Mol Genet 1999;8:237–245
- Kruglyak L. Prospects for whole-genome linkage disequilibrium mapping of common disease genes. Nat Genet 1999;22:139–144
- Murayama S, Saito Y. Neuropathological diagnostic criteria for Alzheimer's disease. Neuropathology 2004;24:254–260
- Braak H, Braak E, Bohl J. Staging of Alzheimer-related cortical destruction. Eur Neurol 1993;33:403–408
- Ahmadian A, Gharizadeh B, Gustafsson AC, et al. Single-nucleotide polymorphism analysis by pyrosequencing. Anal Biochem 2000;280: 103–110
- Lai E, Riley J, Purvis I, et al. A 4-Mb high-density single nucleotide polymorphism-based map around human APOE. Genomics 1998;54: 31–38
- Hao K, Li C, Rosenow C, et al. Detect and adjust for population stratification in population-based association study using genomic control markers: an application of Affymetrix GeneChip Human Mapping 10K array. Eur J Hum Genet 2004;12:1001–1006
- Hua J, Craig DW, Brun M, et al. SNiPer-HD: improved genotype calling accuracy by an expectation-maximization algorithm for high-density SNP arrays. Bioinformatics 2007;23:57–63
- Barrett JC, Fry B, Maller J, et al. Haploview: analysis and visualization of LD and haplotype maps. Bioinformatics 2005;21:263–265
- Corder EH, Saunders AM, Strittmatter WJ, et al. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. Science 1993;261:921–923
- Selkoe DJ, Podlisny MB. Deciphering the genetic basis of Alzheimer's disease. Annu Rev Genomics Hum Genet 2002;3:67–99
- Bertram L, McQueen MB, Mullin K, et al. Systematic meta-analyses of Alzheimer disease genetic association studies: the AlzGene database. Nat Genet 2007;39:17–23

Editor's Note: We encourage authors to submit papers for consideration as a part of our Focus on Alzheimer's Disease and Related Disorders section. Please contact Eric M. Reiman, M.D., at Eric Reiman@bannerhealth.com.